

Coordination of Diatomic Ligands to Heme: Simply CO

Nathan J. Silvernail,[†] Bruce C. Noll,[†] Charles E. Schulz,^{*‡} and W. Robert Scheidt^{*†}*Department of Chemistry and Biochemistry, University of Notre Dame, Notre Dame, Indiana 46556, and Department of Physics, Knox College, Galesburg, Illinois 61401*

Received July 18, 2006

The synthesis and molecular structures of three iron(II) porphyrins with only CO as the axial ligand(s) are reported. Two five-coordinate [Fe(OEP)(CO)] derivatives have Fe–C = 1.7077(13) and 1.7140(10) Å, much shorter than those of six-coordinate [Fe(OEP)(Im)(CO)], although $\nu_{\text{C-O}}$ is 1944–1948 cm^{-1} . The six-coordinate species [Fe(OEP)(CO)₂] has also been studied. The competition for π -back-bonding of two CO ligands leads to Fe–C distance of 1.8558(10) Å and $\nu_{\text{C-O}}$ being increased to 2021 cm^{-1} . The Mössbauer spectrum has a quadrupole splitting constant of 0 mm/s at 4.2 K, indicating high electronic symmetry.

Many heme-based sensing proteins that use/detect the diatomic ligands O₂, CO, or NO are known.^{1–4} These sensing proteins rely on variations in the coordination number and the character of their axial ligands to induce conformational changes leading to protein activation.¹ Effects involving the trans ligand (or lack thereof) on binding diatomic molecules are important in elucidating the mechanism of small-molecule sensing proteins. To achieve a better understanding of the heme interactions involved with diatomic ligand sensing as well as broadly extending our understanding of their electronic and molecular structure, we have been investigating heme/diatomic ligand interactions.

We report for the first time the solid-state syntheses and structural characterization of unambiguously five- and six-coordinate heme CO complexes: [Fe(OEP)(CO)], [Fe(OEP)(CO)]·C₆H₆, and [Fe(OEP)(CO)₂].^{5,6} Although these species have been previously reported in solution and the CO binding

constants determined for three different porphyrins,⁷ we now demonstrate the structural effects of changing the coordination environments. We compare the five-coordinate structures with those of several other diatomic ligand heme complexes, examine the effects of the addition of a sixth ligand, and note structural differences.

These adducts may elucidate important coordination chemistry features involving ligand loss, ligand switching, and ligand photolability. CO photolysis of six-coordinate heme carbonyl derivatives has been a mainstay in biophysical investigations⁸ and is very efficient, with quantum yields nearing unity. The suggestion that five-coordinate heme carbonyls are less photolabile than their six-coordinate counterparts⁹ generates important questions about structural differences.

We first consider the vibrational properties of five-coordinate [Fe(OEP)(CO)]. $\nu_{\text{C-O}}$ in unperturbed imidazole-ligated hemes is typically near 1970 cm^{-1} , but solid-state environmental factors can lead to variation in $\nu_{\text{C-O}}$ in the range of 1926–2000 cm^{-1} .^{10,11} Thus, it is perhaps surprising to find that $\nu_{\text{C-O}}$ in [Fe(OEP)(CO)] and [Fe(OEP)(CO)]·C₆H₆ is well within this range at 1944 and 1948 cm^{-1} , respectively (Nujol mull). These values might suggest that the Fe center is involved in a significant intermolecular interaction to form a pseudo-six-coordinate complex, but this is not correct (vide infra).

The molecular structures of the two crystalline [Fe(OEP)(CO)] complexes are illustrated in Figure 1 (and Figure S1 in the Supporting Information). There are strong similarities as well as differences in their structures: the Fe out-of-plane displacements are similar at 0.20 or 0.22 Å, the off-axis tilt of the Fe–C vector is 3.8° or 2.4°, and the Fe–C–O bond angle is 177.20(8)° in both. Both forms show some ring–

* To whom correspondence should be addressed. E-mail: (C.E.S.), Scheidt.1@nd.edu (W.R.S.).

[†] University of Notre Dame.

[‡] Knox College.

- (1) Gilles-Gonzalez, M.-A.; Gonzalez, G. *J. Inorg. Biochem.* **2005**, *99*, 1.
- (2) Rodgers, K. R.; Lukat-Rodgers, G. S. *J. Inorg. Biochem.* **2005**, *99*, 963.
- (3) (a) Roberts, G. P.; Kerby, R. L.; Youn, H.; Conrad, M. *J. Inorg. Biochem.* **2005**, *99*, 280. (b) Aono, S.; Nakajima, H. *Coord. Chem. Rev.* **1999**, *190–192*, 267.
- (4) Boon, E. M.; Marletta, M. A. *J. Inorg. Biochem.* **2005**, *99*, 892.
- (5) The following abbreviations are used in this paper: Por, generalized porphyrin dianion; OEP, dianion of octaethylporphyrin; Deut, dianion of deuteroporphyrin; TPP, dianion of tetraphenylporphyrin.
- (6) Complete synthetic procedures are given in the Supporting Information.

- (7) (a) Rougee, M.; Brault, D. *Biochemistry* **1975**, *14*, 4100. (b) Wayland, B. B.; Mehne, L. F.; Swartz, J. *J. Am. Chem. Soc.* **1978**, *100*, 2379. (c) Strauss, S. H.; Holm, R. H. *Inorg. Chem.* **1982**, *21*, 863.
- (8) Sage, J. T.; Champion, P. M. Small Substrate Recognition in Heme Proteins. In *Comprehensive Supramolecular Chemistry*; Suslick, K. S., Ed.; Pergamon: Oxford, U.K., 1996; Vol. 5, pp 171–217.
- (9) (a) Pal, B.; Li, Z.; Ohta, T.; Takenaka, S.; Tsuyama, S.; Kitagawa, T. *J. Inorg. Biochem.* **2004**, *98*, 824. (b) Makino, R.; Obayashi, E.; Homma, N.; Shiro, Y.; Hori, H. *J. Biol. Chem.* **2003**, *278*, 11130.
- (10) Silvernail, N. J.; Roth, A.; Schulz, C. E.; Noll, B. C.; Scheidt, W. R. *J. Am. Chem. Soc.* **2005**, *127*, 14422.
- (11) Kim, K.; Ibers, J. A. *J. Am. Chem. Soc.* **1991**, *113*, 6077.

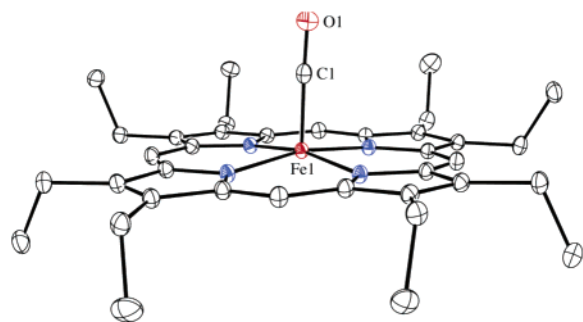


Figure 1. Thermal ellipsoid plot of [Fe(OEP)(CO)] (50% probability ellipsoids). H atoms are omitted for clarity.

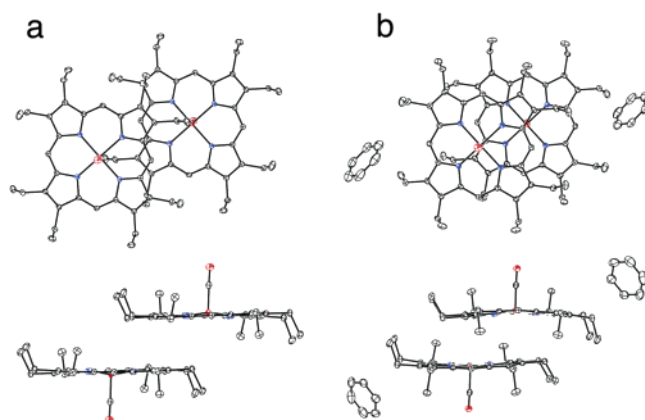


Figure 2. ORTEP plots (50% probability ellipsoids) of [Fe(OEP)(CO)] (a) and [Fe(OEP)(CO)]·C₆H₆ (b) displaying the pairwise interactions. In part a, the 24-atom mean-plane separation is 3.42 Å and the lateral shift of the two ring centers is 6.76 Å. The Fe···Fe distance is 7.58 Å. The corresponding distances in part b are 3.46, 3.88, and 5.20 Å, respectively.

ring interactions, although these lead to some strong differences as well. Figure 2a shows the pairwise interactions in [Fe(OEP)(CO)], while the equivalent packing in the benzene solvate is shown in Figure 2b. In both cases, there is inversion symmetry; metrical information is given in the figure caption. The inter-ring packing pattern in the unsolvated form of [Fe(OEP)(CO)] (Figure 2a) is similar to that of both crystalline forms of [Fe(OEP)(NO)], for which little porphyrin overlap is observed.¹² The closest intermolecular contact to the Fe in [Fe(OEP)(CO)] is 3.12 Å, whereas in [Fe(OEP)(CO)]·C₆H₆, the closest contact to Fe is 3.58 Å. In the two [Fe(OEP)(NO)]⁺ structures (isoelectronic to the CO's), however, there is a large porphyrin ring overlap with the π system of an adjacent molecule acting as a pseudo sixth ligand.¹³ The tighter interactions in [Fe(OEP)(CO)]·C₆H₆ (Figure 2b) are comparable to, although slightly larger than, those in the isoelectronic [Fe(OEP)(NO)]⁺ complexes. The porphyrin core conformation in [Fe(OEP)(CO)] is nearly planar, whereas that in [Fe(OEP)(CO)]·C₆H₆ displays modest core ruffling. The average equatorial Fe–N_p distances are 1.988 (2) and 1.984(3) Å for the unsolvated and solvated forms, respectively. The very short Fe–N_p bond distances reflect the strong bonding interaction and low-spin state of Fe.

Significantly, the axial-ligand-induced equatorial (Fe–N_p) bond distance differences observed in five-coordinate [Fe(por)(NO)] derivatives¹² are not observed; individual Fe–N_p distances are tightly clustered.

The short axial Fe–C distance of 1.7140(11) Å in [Fe(OEP)(CO)] is shorter than those of the six-coordinate imidazole adducts by ~ 0.03 – 0.06 Å.^{10,14} This might suggest stronger Fe \rightarrow C π back-donation, but the relatively normal ν_{C-O} value suggests that the Fe π donation into the CO π^* orbitals is similar to that of the six-coordinate species. This would then suggest that it is the σ -bonding component that leads to the shorter Fe–C bond distance. The distance in [Fe(OEP)(CO)]·C₆H₆ is 1.7077(13) Å; the possibly slightly shorter Fe–C distance is not consistent with the 4-cm⁻¹ increase in ν_{C-O} , again suggesting the importance of σ bonding. We thus conclude that differences in the σ -bonding component of Fe–C have no or modest effects on ν_{C-O} . As we have noted for a different series of six-coordinate carbonyl derivatives,¹⁰ there is a strong correlation between ν_{C-O} and the Fe–C/C–O bond lengths; this question is being examined in more detail for OEP derivatives.

The structures of other [Fe(OEP)(XY)]⁺⁰ adducts (XY = NO or CS) are available for comparison. Structural parameters for five- and six-coordinate species are listed in Table 1. The Fe out-of-plane displacement of [Fe(OEP)(CO)] is seen to be at the low end of the range. In all cases, the addition of a sixth ligand leads to a decreased Fe atom displacement and in most cases to an increase in the length of the Fe–X(XY) bond. Although this increase is never large, the CO's are the system that experiences the largest increase in the Fe–C bond distance {from 1.7140(11) to 1.7733(12) Å in [Fe(OEP)(CO)(1-MeIm)]}.¹⁴ Interestingly, the Fe–C bond would appear to have changed minimally when the trans ligand is the weakly coordinating tetrahydrofuran (THF) ligand in [Fe(Deut)(CO)(THF)]. Unfortunately, the relatively low precision of this structure (from ~ 25 years ago) is not adequate to comment on further but clearly merits further attention.

Although solutions of Fe(OEP) and CO will always be mixtures of mono- and bis-CO species, crystallization experiments (see the Supporting Information) at low temperature reproducibly afford crystals of [Fe(OEP)(CO)₂]. Analysis of crystals of [Fe(OEP)(CO)₂] gave the structure displayed in Figure 3; the complex has a required inversion center. As might be expected, the competition from two CO's for π back-bonding from the central Fe leads to increased Fe–C bond distances of 1.8558 (10) Å, which is also consistent with the observed asymmetric stretch of 2021 cm⁻¹. The CO ligands are tilted off-axis as shown with other metrical information given in Figure 3. Perhaps as a consequence of the competition for bonding with two CO ligands, the equatorial Fe–N_p bonds are at the very long end of values expected for low-spin Fe^{II}.¹⁵ The porphyrin

(12) (a) Ellison, M. K.; Scheidt, W. R. *J. Am. Chem. Soc.* **1997**, *119*, 7404. (b) Scheidt, W. R.; Duval, H. F.; Neal, T. J.; Ellison, M. K. *J. Am. Chem. Soc.* **2000**, *122*, 4651.
(13) Ellison, M. K.; Schulz, C. E.; Scheidt, W. R. *Inorg. Chem.* **2000**, *39*, 5102.

(14) Silvermail, N. J.; Noll, B. C.; Scheidt, W. R., unpublished results.
(15) Scheidt, W. R. Systematics of the Stereochemistry of Porphyrins and Metalloporphyrins. In *The Porphyrin Handbook*; Kadish, K. M., Smith, K., Guillard, R., Eds.; Academic Press: San Diego, CA, and Burlington, MA, 2000; Vol. 3, Chapter 16.

Table 1. Notable Bonding Parameters for [Fe(OEP)(CO)] and Related Compounds

complex	Fe–XY ^a	X–Y ^a	Fe–X–Y ^b	Fe–Np ^a	Fe–L _{ax} ^a	ΔFe ^c	ν _{X–Y} ^d	ref
[Fe(OEP)(CO)]	1.7140(11)	1.1463(12)	177.20(8)	1.988(2)		0.20	1944 ^e	tw
[Fe(OEP)(CO)]·C ₆ H ₆	1.7077(13)	1.1259(16)	177.20(11)	1.984(3)		0.22	1948 ^e	tw
[Fe(OEP)(NO)]	1.722(2)	1.167(3)	144.4(2)	2.004(15)		0.29	1666 ^e	12
[Fe(OEP)(NO)]	1.7307(7)	1.1677(11)	142.74(8)	2.010(13)		0.27	1673 ^e	12
[Fe(OEP)(NO)] ⁺	1.6528(13)	1.140(2)	173.19(13)	1.994(5)		0.32	1838 ^e	13
[Fe(OEP)(NO)] ⁺	1.644(3)	1.112(4)	176.9(3)	1.994(1)		0.29	1868 ^e	16
[Fe(OEP)(CS)]	1.662(3)	1.559(3)	176.3(2)	1.982(5)		0.23	1292 ^e	17
[Fe(Deut)(CO)(THF)]	1.706(5)	1.144(5)	178.3(14)	1.98(3)	2.127(4)	0.10	1955 ^f	18
[Fe(OEP)(CO)(1-MeIm)]	1.744(5)	1.158(5)	175.1(4)	2.000(3)	2.077(3)	0.00	1965 ^g	19
[Fe(OEP)(CO)(1-MeIm)]	1.7733(12)	1.1413(15)	175.67(11)	2.010(4)	2.0544(9)	0.02	1980 ^e	20
[Fe(OEP)(NO)(1-MeIm)] ⁺	1.6465(17)	1.135(2)	177.28(17)	2.003(5)	1.9889(16)	0.02	1921 ^e	21
[Fe(TPP)(NO)(1-MeIm)]	1.750(2)	1.182(3)	137.7(2)	2.008(13)	2.173(2)	0.04	1628 ^e	22
[Fe(OEP)(CS)(1-MeIm)]	1.703(4)	1.563(4)	172.2(2)	2.001(4)	2.112(3)	0.10	1272 ^e	23
[Fe(OEP)(CO) ₂]	1.8558(10)	1.1216(13)	173.95(9)	2.0133(7)	1.8558(10)	0.00	2021 ^e	tw

^a In angstroms. ^b In degrees. ^c Displacement from the 24-atom mean plane. ^d In reciprocal centimeters. ^e Nujol mull. ^f THF. ^g CD₂Cl₂ solution.

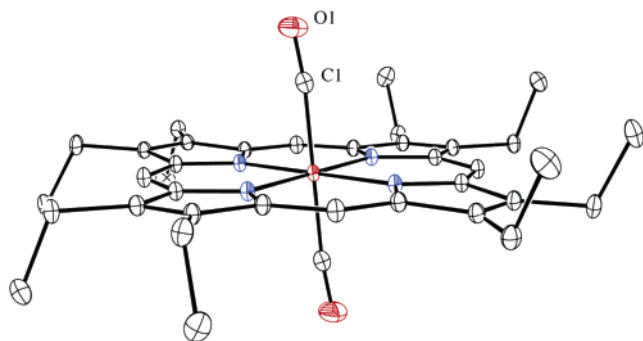


Figure 3. Thermal ellipsoid plot of [Fe(OEP)(CO)₂]. The Fe–C vector is tilted from the heme normal by 5.9°; the C and O atoms are shifted laterally by 0.14 and 0.56 Å, respectively.

core is planar; core diagrams for all three complexes are given in Figure S2 in the Supporting Information.

Mössbauer spectra for [Fe(OEP)(CO)] are consistent with an isolated five-coordinate Fe center. The quadrupole splitting is much larger (1.84 mm/s, 4.2 K) than that observed for six-coordinate carbonyls (typically less than 0.7 mm/s), strongly indicative of a d-orbital asymmetry consistent with five-coordination. The isomer shift value of 0.27 mm/s is similar to that of the six-coordinate derivatives; the relatively low value is consistent with strong and effectively equivalent covalency among the occupied Fe 3d orbitals in all derivatives. The six-coordinate derivative is much more symmetric, with a quadrupole splitting of 0 or near 0 at 4.2 K and an isomer shift of 0.31 mm/s; this increases to a quadrupole splitting of 0.176 mm/s and an isomer shift of 0.18 mm/s at 298 K. Spectra in the applied magnetic field for this complex confirmed that it is a diamagnetic species. Complete data are given in Table S1 in the Supporting Information.

Five-coordinate carbonyl hemes have been structurally characterized and compared with related five- and six-coordinate diatomic complexes. Although the biological importance of five-coordinate carbonyl hemes is not known, proteins have been described that have spectroscopic properties that may be associated with this adduct.^{9,24} Additional spectroscopic and photophysical studies on these carbonyl complexes are in prospect.

Acknowledgment. We thank the NIH for support (Grant GM-38401 to W.R.S.) and the NSF for X-ray instrumentation (Grant CHE-0443233). We thank Prof. Timothy Sage for useful discussions.

Supporting Information Available: Syntheses, figures, and crystallographic data are available (including CIF files). This material is available free of charge via the Internet at <http://pubs.acs.org>.

IC0613356

- Scheidt, W. R.; Lee, Y. J.; Hatano, K. *J. Am. Chem. Soc.* **1984**, *106*, 3191.
- Scheidt, W. R.; Geiger, D. K. *Inorg. Chem.* **1982**, *21*, 2056.
- Scheidt, W. R.; Haller, K. J.; Fons, M.; Mashiko, T.; Reed, C. A. *Biochemistry* **1981**, *20*, 3653.
- Salzmann, R.; McMahon, M. T.; Godbout, N.; Sanders, L. K.; Wojdelski, M.; Oldfield, E. *J. Am. Chem. Soc.* **1999**, *121*, 3818.
- Silvermail, N. J.; Noll, B. C.; Scheidt, W. R., unpublished results.
- Ellison, M. K.; Scheidt, W. R. *J. Am. Chem. Soc.* **2001**, *123*, 5210.
- Wyllie, G. R. A.; Schulz, C. E.; Scheidt, W. R. *Inorg. Chem.* **2003**, *42*, 5722.
- Cao, C.; Dahal, S.; Shang, M.; Beatty, A. M.; Hibbs, W.; Schulz, C. E.; Scheidt, W. R. *Inorg. Chem.* **2003**, *42*, 5202.
- (a) Chu, G. C.; Katakura, K.; Tomita, T.; Zhang, X.; Sun, D.; Sato, M.; Sasahara, M.; Kayama, T.; Ikeda-Saito, M.; Yoshida, T. *J. Biol. Chem.* **2000**, *275*, 17494. (b) Vogel, K. M.; Spiro, T. G.; Shelver, D.; Thorsteinsson, M. V.; Roberts, G. P. *Biochemistry* **1999**, *38*, 2679.